**Neuroprotective effect of secretin in chronic hypoxia induced neurodegeneration in rats**

Gowtham Padmanaban¹*, M. K. Kayalvizhi¹, Kalyanasundaram Kasiviswanathan², Ruckmani Arunachalam², Vishnu Kumar Urkavalan³

1Department of Pharmacology, Adhiparaskathi Institute of Medical Sciences, Melmaruvathur, Tamilnadu, India
2Department of Pharmacology, Chettinad Hospital and Research Institute, Tamilnadu, India
3AMR Pharma Pvt Ltd, Chennai, Tamil Nadu, India

**ABSTRACT**

**Background:** Hypoxia is a condition in any stage in the delivery of oxygen to cells which include decreased partial pressures of oxygen, less diffusion of oxygen in the lungs, insufficient hemoglobin, inefficient blood flow to the end tissue, and breathing rhythm. Secretin is an amino acid which plays proper functioning of gastro intestinal system.

**Methods:** The current study was conducted to evaluate the effect of exogenously administrated secretin on chronic hypoxic damage of brain in rat model. Experimental design consists of control animals, Control animals + secretin hypoxia exposed animals; hypoxia exposed animals +secretin (20ng/kg.bw).

**Results:** The results of this study point to a possible role of Secretin as neuroprotectant.

**Conclusions:** Further research on secretin needs to be conducted in order to confirm the deductions made by this study.

**Keywords:** Anti-oxidant, Hypoxia, Secretin, Hemoglobin, Neuroprotectant

**INTRODUCTION**

Hypoxia a condition in which the level of oxygen supplied to the body/tissue becomes inadequate. The need for adequate oxygen and glucose supply as well as removal of carbon dioxide is essential for tissue homeostasis. Hypoxia can result from a failure at any stage in the delivery of oxygen to cells which include decreased partial pressures of oxygen, less diffusion of oxygen in the lungs, insufficient hemoglobin, inefficient blood flow to the end tissue, and breathing rhythm. There are four basic causes for hypoxia viz, anoxic hypoxia or diffusion hypoxia, anaemic hypoxia, stagnant hypoxia and histotoxic (histologic) hypoxia.

Hypoxias can occure both acute and chronic. Acute hypoxia is a sudden or rapid depletion in available oxygen at the tissue level. The condition may result from asphyxia, airway obstruction, acute hemorrhage, blockage of alveoli by edema or infectious exudate, or abrupt cardiorespiratory failure. Chronic hypoxia is a
usually slow, insidious reduction in tissue oxygenation resulting from gradually destructive or fibrotic lung diseases, congenital or acquired heart disorders, or chronic blood loss.

The symptoms of Hypoxia includes, severity and duration like altitude sickness, where hypoxia develops gradually, the symptoms include light-headedness, fatigue, numbness, tingling of extremities, nausea and anorexia.1

Secretin is initially synthesized as a 120 amino acid precursor protein known as prosecretin. The mature secretin peptide is a linear peptide hormone, which is composed of 27 amino acids and has a molecular weight of 3055. The amino acids sequences of secretin have some similarities to that of glucagon, vasoactive intestinal peptide (VIP), and gastric inhibitory peptide (GIP).

Fourteen of 27 amino acids of secretin reside in the same positions as in glucagon, 7 the same as in VIP, and 10 the same as in GIP. The principle action of secretin is to stimulate bicarbonate secretion to neutralize gastric acid in the duodenum mediated by the secretin receptor, a G-protein coupled secretin receptor. The receptor is structurally similar to receptors for VIP, glucagon, parathyroid hormone and other Class II G-protein linked receptors. Secretin receptors have been identified by ligand binding and recognition of its mRNA in pancreas, biliary system, stomach, brain and kidney.

Several research works was performed with secretin. When secretin was injected, it decreased the approaches to novel items, movement in an open environment and respiration in rats. Secretin signal plays a neuroprotective role against the neurotoxicity of ethanol induced developmental neurodegeneration in secretin receptor-deficient mice.4 And some of the research works observed that increased concentrations of glutamate and GABA in rat hippocampus following secretin application. Secretin depressed the effects of single doses of morphine in mice and increased the delay before the animals jumped to avoid an aversive stimulus.5 The role secretin in brain metabolism during hypoxia was reported byyungetal. Till a decade ago secretin was known for its gastro intestinal fuctions. But it receptors have been defined and its pleiotropic effects outside GIT has been reported by many authors. As secretin has pleiotropic effect, it was planned to study its action in the hypoxia in rats.

METHODS

Secretin was purchased from Phoenix Pharmaceuticals, USA and all other chemical used were of analytical grade obtained from sisco research laboratory, Mumbai, India. Animal experiments were carried out after getting clearance from the Institutional Animal Ethical Committee (IAEC No: 06/July 13). The experimental animals were healthy, inbred 20 adult male Wistar albino rats, weighing between 200 - 220g (12 weeks of age) were used. The animals were maintained under standard laboratory conditions and allowed to have food and water. All the rats were housed under conditions of controlled temperature (26±2 °C) with 12hr light and 12hr dark exposure.

Hypoxic chamber

Hypoxic chamber consists of an air tight box made up of acriline material in which provision was made for the administration of hypoxic gas. Hypoxic gas was a combination of 92% nitrogen and 8% oxygen delivered through a separate cylinder (B type 10.0 litres). Gas cylinder was purchased from TN oxygen Pvt. Ltd, Ambuttar.

Method

Experimental design consists of Group I - was control animals (normoxia); Group II- Control animals + secretin (20ng/kg.bw); Group III - hypoxia exposed animals; Group IV - hypoxia exposed animals +secretin (20ng/kg.bw). Animals of Group III and Group IV were exposed to hypoxia, by keeping the animals in hypoxic chamber connected to a gas flow of 92% nitrogen and 8% oxygen for 45 minutes each day for 14 days.6 On the final day of hypoxia exposure, Group II and Group IV were administrated with Secretin intraperitoneally (20 ng/BW) once daily for 3 consecutive days.

Sample collection

On the 17th day hippocampus was isolated after sacrificing the animals under deep anesthesia using Halothene. Hippocampus was excised, washed in ice cold saline and blotted to dryness. The hippocampus samples were homogenized by using Teflon glass homogenizers. 10% homogenate hippocampus of tissue was prepared in phosphate buffer (0.1 M, pH 7.0) and centrifuged at 3000 rpm at 4°C for 15 min to remove cell debris and the clear supernatant was used for further biochemical assays.7 The results are expressed as mean ± standard deviation (SD). All data were analyzed with the SPSS for windows statistical package (version 20.0, SPSS Institute Inc., Cary, North Carolina. Statistical significance between the different groups was determined by one way-analysis of variance (ANOVA). When the groups showed significant difference then Tukey’s multiple comparison tests was followed and the significance level was fixed at p< 0.05.

I. Biochemical determinations

Assay of lipid peroxidation
Lipid peroxidation (LPO) was determined and the Malondialdehyde (MDA) forms as an intermediate product of the peroxidation of lipids and serves as an index of the intensity of oxidative stress. MDA reacts with thiobarbituric acid to generate a colored product which absorbs light at 532 nm.

Nitric oxide analysis

Nitric oxide (NO) levels were measured as total nitrite + nitrate levels by the Griess reagent consists of sulfanilamide and $N$-(1-naphthyl)-ethylenediamine. The method is based on a two-step process. The first step is the conversion of nitrate into nitrite using a nitrate reductase. The second step is the addition of the Griess reagent, which converts nitrite into a deep purple azo compound; photometric measurement of absorbance at 540 nm is due to the fact that this azochromophore accurately determines nitrite concentration.

Superoxide Dismutase (SOD) assay

Superoxide dismutase (SOD) was analysed and Pyrogallol auto oxidizes rapidly in an aqueous solution at a fast rate with higher (pH=8.0) to produce several intermediate products. The inhibitions of pyrogallol auto oxidation by the enzyme present in sample is employed in the quantification of activity of SOD. The inhibition of auto oxidation brought about by the addition of enzyme, which is evaluated at the early stage as an increase in absorbance at 420 nm in 0, 1, 2 and 3 min intervals. The unit of enzyme activity is defined as the enzyme required for 50% inhibition of pyrogallolautooxidation.

Catalase (CAT) assay

The activity of Catalase (CAT) was analysed, dichromate in acetic acid is reduced to chromic acetate, when heated in the presence of hydrogen peroxide ($H_2O_2$) perchloric acid formed as an unstable intermediate. Since dichromate has no absorbance in this region. The catalase preparation is allowed to split $H_2O_2$ for different period of time and the reaction was arrested by the addition of dichromate/acetic acid mixture at a particular interval of 0, 15, 30 and 60 s. The remaining $H_2O_2$ is determined by measuring chromic acetate at 610 nm.

Glutathione peroxidase (GPx) assay

Glutathione peroxidase (GPx) activity was estimated by the reaction between glutathione remaining after the action of GPx and 5, 5'-dithiobis-(2-nitrobenzoic acid) (DTNB) to give a compound that absorbs light at 412 nm.

II. Estimation of non-enzymatic antioxidants

Glutathione (GSH) analysis

Reduced glutathione (GSH) in the hippocampus were estimated by the reaction of GSH with DTNB that gives a compound which absorbs light at 412 nm.

Ascorbic acid (Vitamin - C) assay

Ascorbic acid was assayed by reacting with copper to form dehydroascorbic acid and diketogluutaric acid, which was then treated with 2,4-dinitrophenyl hydrazine to form the derivative of bis-2,4-dinitrophenyl hydrazine. This compound in sulphuric acid undergoes a rearrangement to form a product which absorbs light at 520 nm.

RESULTS

Biochemical assays

The data are presented with Mean ±SD in (Figures 1 to 7). There were no significant differences in antioxidant assays between control and secretin control animals. In hypoxia exposed animals LPO, SOD, CAT, Gpx, GSH and Vitamin C were significantly increased when compared to control and secretin control animal. However hypoxia animals treated with secretin were significantly decreased when compare to hypoxia exposed animals.
DISCUSSION

The current study was conducted to evaluate the effect of exogenously administrated secretin on chronic hypoxic damage of brain. Chronic hypoxia resulted in impairing the novelty-seeking behaviour and working memory. Chronic hypoxia produces morphological changes in the hippocampus and causes change in the band appearance of CA 1. These morphological changes may make the hippocampus vulnerable to damage by metabolic challenges, and prolonged oxidative stress can lead to cell death. These chronic hypoxia -induced morphological changes lead to deficits in working memory. The measures of working memory clearly suggest that chronic hypoxia is detrimental to working memory. The experiment reveals a significant induction of lipid peroxidation in the rat brain homogenate of hypoxic groups. Enhanced lipid peroxidation is associated with stimulation of arachidonic acid metabolism and prostaglandin formation. A close association among phospholipid methylation, arachidonic acid release from phospholipid and increased prostaglandin was found. The protective effects of Secretin on hypoxia induced lipid peroxidation (Figure 7) could be directly attributed to its antioxidant property. As mentioned earlier, hypoxia has been postulated to facilitate the production of arachidonic acid, which could contribute to the oxidative stress in the form of lipid peroxidation. Thus Secretin could be reducing hypoxia - induced increase in arachidonic acid production. Superoxide toxicity appears to be through an indirect action on living cells by giving rise to more powerful oxidants such as the hydroxyl radical. The dismutation of O$_2^-$ generates H$_2$O$_2$ and oxygen, and this reaction is a spontaneous reaction which can also be catalysed by the enzyme superoxide dismutase. Secretin decreases superoxide dismutase, making cell to clear the free radicals. Hence, superoxide dismutase increases when free radicals increase. In hypoxia treated group superoxide dismutase levels were high compared to other. This shows that hypoxia can elevate free radicals to high level, whereas in hypoxic + secretin treated groups, Superoxide dismutase level is decreased compared to hypoxic group. Hence Secretin acts as an antioxidant by decreasing the free radicals.
Traumatic injury of the brain is characterised by disruption of cell bodies and axons, followed by little or no axonal regeneration and virtually no recovery of function by the lesioned tissue. These results clearly demonstrate the structural damage that hypoxia causes in the CA1 region of the hippocampus, the part of the brain responsible for learning and memory processing.

The rise in intracellular levels of calcium following hypoxic treatment may be due to either hypoxic stimulating calcium entry or hypoxic inhibiting calcium exit and storage. The increased swelling and necrosis of the cells in the CA1 region may be attributable to increased intracellular levels of calcium as a direct consequence of hypoxia exposure. In hypoxic exposed + secretin treated groups, the cells of CA1 band is slightly disturbed may due to hypoxic stress, but nucleus ,cell membrane are clear. Hence, neurons are normal in spite of hypoxic stress, this may due to protective effect of Secretin. T.

The results of this study point to a possible role of Secretin as neuroprotectant, further research on secretin needs to be conducted in order to confirm the deductions made by this study.

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REFERENCES
